

INSTRUCTIONS FOR USE

MPB004

µPREP™ Half Fraser Broth ISO (+FAC)

A sterile, ready to reconstitute primary enrichment medium for the isolation of *Listeria* spp. from food samples.

Presentation: This kit contains 5 sterile single use µPREP™ bags.

Contents

µPREP™ Half Fraser Broth ISO (+FAC) 20 litre bag(s), sterile in outer bags.

Each µPREP™ Bag is supplied with the following attachments:

- Tube
- Slide clamp
- Black connector and cap

Materials required but not provided

1. MPA001 µPREP™ Filter Unit
2. MPA002 µPREP™ Quick Connectors
3. Sterile tubing
4. Reverse Osmosis (RO) or deionised water source
5. Peristaltic pump
6. Dispensing system

Composition

	Grams per litre
Peptone mixture	15.0
Yeast extract	5.0
Aesculin	1.0
Disodium hydrogen phosphate	9.6
Potassium dihydrogen phosphate	1.35
Sodium chloride	20.0
Lithium chloride	3.0
Acridine	0.0125
Nalidixic acid	0.01
Ferric ammonium citrate	0.5

Final pH: 7.2 ± 0.2

Hazard classification: NR – Not regulated

Appearance: reconstituted liquid should be tan with good clarity

Storage:

Bag (as supplied): store in the dark at 10-25°C

Bag (reconstituted): store in the dark at 10-25°C for up to 72 hours (providing asepsis is maintained)

Minimum Q.C. organisms

Listeria monocytogenes ATCC 13932

Escherichia coli ATCC 25922

Enterococcus faecalis ATCC 29212

Product Performance and Limitations

Performance is guaranteed as long as manufacturer's guidelines are followed. Aseptic technique must be observed during reconstitution and use.

Restrictions on Use

This product is sold for use only in the following fields: food, animal feed, beverages, pharmaceuticals or environmental. This product is not sold for use with human clinical or veterinary material and is not designed for human or therapeutic procedures.

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Suggested protocol for use of Lab M µPREP™ HFB ISO (+FAC) Bag for 20 litres

1. Prepare a sterile MPA001 µPREP™ Filter Unit, ensuring any kinks in the tubing are removed. Remove µPREP™ bag and attachments carefully from the outer bag. Unfold the µPREP™ bag.
2. Aseptically remove both the white cap from the white MPA001 µPREP™ Filter Unit tubing connector and the black cap from the black µPREP™ bag tubing connector. Retain both caps aseptically for later use.
3. Attach the white MPA001 Filter Unit connector to the black µPREP™ bag connector.
4. Prepare a tube, running from a 20 litre RO/deionised water source via peristaltic pump. For best results, prime the tubing to remove excess air before connecting to the filter unit via the barb (push on connection). Prime the pump on a slow setting initially to allow time for adequate removal of trapped air (step 5).
5. Before activating the pump to commence filling, release the slide clamp on the µPREP™ bag tubing and note:

for optimum filter performance, it is recommended that the filter be held at a 45° angle (with screw valve to be in the uppermost position) thus allowing full expulsion of air from the device. To prevent build up of pressure upon commencement of filling, slowly turn the screw valve on the side of the capsule filter to release trapped air from the filter unit. Do NOT fully remove the screw. N.B. Purging air and filling of filter unit with water happens rapidly. Ensure the screw is securely tightened after releasing the air to avoid water leaks.

The speed of the peristaltic pump may then be increased.

6. During filling, manipulate the bag to aid dissolution of powder. Tubing should be monitored for build up of pressure which can be identified by the tubing between the peristaltic pump and filter becoming rigid. Ensure the water supply does not run dry during filling – any air pumped into the filter unit will cause a build up of pressure within the connected systems. Care should be taken not to fill the µPREP™ bag with more than 20 litres of water.
7. Once the bag has been filled with 20 litre RO/deionised water, turn off the pump. To drain any remaining water from the tubing into the bag, it may be necessary to gently squeeze the bag to force air into the tubing, alternatively, the tubing may be elevated and squeezed. The slide clamp should then be applied. It is recommended that the slide clamp be applied near to the bag rather than connector end of the tubing. Check the integrity of the bag and ensure powder is fully dissolved. At this point, the connectors should be unscrewed and the bag cap can be reapplied or the bag connector can be immediately connected to the dispensing system (step 9).
8. Remove the filter unit and reattach the cap. The MPA001 µPREP™ Filter Unit may be autoclaved and reused; record the usage of the filter and discard after 100 litres or 5 filled bags. If the flow rate is greatly reduced at any time, the filter should be discarded and replaced.
9. Attach a sterilised quick connector (supplied separately) and sterilised tubing set to the bag tubing via the bag connector.
10. Connect the new tubing to a dispensing system or peristaltic pump, open the slide clamp and begin dispensing the reconstituted, sterile culture media as required. At low volumes it may be necessary to manipulate the bag to enable full drainage. Care should be taken not to allow air to be taken into system as this will adversely affect accuracy of dispensing.
11. When dispensing is complete, disconnect bag tubing from dispensing tube. Retain the quick connector and cap and sterilise for future use.
12. If the µPREP™ Bag is not emptied and media is required to be stored, close the slide clamp and use the sterile (if asepsis has been maintained) cap to reseal the bag tubing. Once the µPREP™ Bag is emptied, dispose with normal laboratory waste.

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