PRIOR TO STARTING THE TEST:

Note: Food Samples should be prepared and enriched, following the directions outlined in the GeneQuence Salmonella spp. test kit insert.

UPON OPENING THE KIT:

1. Add 6 mL of lysis reagent buffer (bottle 1b) directly to lysis reagent concentrate (bottle 1a).
2. Mix wash solution (bottle 4) with 950 mL of distilled or deionized water.
3. For each sample and control, label a 12 x 75 mm glass tube with appropriate sample designation.
4. Add 0.1 mL of reconstituted lysis reagent (bottle 1a) to each tube. Incubate the rack of tubes in the 65°C water bath or heater block for 5 minutes.
5. Transfer 0.150 mL of each lysed sample, including the controls, to designated microwells.
6. Add 0.125 mL of hybridization/probe mixture to each microwell, with the exception of the reagent blank microwell. Incubate the plate at 45°C for 60 minutes.
7. Wash the microwells 5 times.
8. Add 0.150 mL of substrate chromagen solution (bottle 5) to each microwell, including the blank microwell. Incubate the plate at room temperature for 20 minutes.
9. Add 0.050 mL of stop solution (bottle 6) to each microwell, including the blank microwell. Read absorbance at 450 nm using a plate strip reader.

TEST PROCEDURE:

If using the 48-hour enrichment, add 0.2 mL of each of the two GN cultures for each sample to the appropriate tubes (0.2 mL of each of the two GN cultures pooled into one tube for each sample and 0.4 mL of each control). If using the 24-hour enrichment, add 0.4 mL of the samples and each control to the appropriately labeled tubes.

While the rack is incubating, refer to chart and mix hybridization solution (bottle 2) and probe solution (bottle 3) in a plastic or glass vial. Mix thoroughly.